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# Synthesis and Antimicrobial Activity of Tetrodecamycin Partial Structures

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Abstract—An efficient synthetic approach to the core structure 5 of the novel polyketide antibiotic tetrodecamycin (1) was developed. This approach features the acid-catalyzed cyclization of a *tert*-butyldimethylsilyl protected methyl α-(γ-hydroxyacyl) tetronate, leading to the novel tricyclic ring skeleton exhibited by 5, and an efficient strategy for the parallel introduction of the *cis*-diol and *exo*-methylene function. In addition to 5, diastereomer 26, analogue 6 and several derivatives (16, 27–29) were prepared and evaluated for their antibacterial activities against *Staphylococcus aureus* (including MRSA) and *Enterococcus faecalis* and for their cytotoxic activities against human leukemia cell lines (HL-60, Jurkat T-cells). While compound 5 did not inhibit the growth of the Gram-positive pathogens (MICs > 128 μg mL<sup>-1</sup>), analogue 6 and 2-naphthoyl derivative 27 showed promising antibacterial activities with MICs of 4–16 μg mL<sup>-1</sup>. Remarkably, the antibacterial activity of these compounds was paralleled by cytotoxicity (IC<sub>50</sub> 10–23 μM). The reactive *exo*-methylene moiety was shown to be crucial, but not sufficient by its own, for both the antibacterial and the cytotoxic activities.

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#### Introduction

New classes of antibacterial agents with novel mechanisms of action are urgently needed, due to the worldwide emergence of infections caused by multidrug resistant bacteria. In particular, resistances to Gram-positive bacteria such as methicillin-resistant *Staphylococcus aureus* (MRSA), methicillin-resistant coagulase negative staphylococci (MRCNS) as well as vancomycin-resistant *Enterococcus faecalis* and *E. faecium* (VRE) give rise to significant problems amongst hospitalized patients.<sup>2</sup>

Tetrodecamycin (1), a novel polyketide antibiotic isolated from the culture broth of *Streptomyces nashvillensis* MJ885-mF8, has been shown to have promising antibacterial activity against Gram-positive bacteria including

Staphylococcus aureus (MIC 6.25–12.5 µg mL<sup>-1</sup>) as well as *Bacillus anthracis*.<sup>3</sup> Markedly enhanced activity was observed for several 14-O substituted derivatives (e.g., 2-naphthoyl derivative 2 with a MIC of 0.78–3.12 µg mL<sup>-1</sup> against *S. aureus* in vitro).<sup>4</sup> On the other hand, dihydrotetrodecamycin (3), which has been isolated from the Streptomyces strain, too, is virtually inactive, revealing the crucial role of the *exo*-methylene moiety in 1 and 2 for biological activity.

Although the mechanism of action of these antibiotics is still unknown, one can speculate, that the biological activities of 1 and 2 may be attributed to their reactivity with the cystein residues of functional proteins forming covalent bonds via Michael type addition. Indeed, thiols like *N*-acetylcysteamine were shown to add smoothly across the *exo*-methylene moiety of 1, leading to the respective *S*-alkylation products (e.g., 4).<sup>4</sup> However, unlike 3, these derivatives still displayed marked antibacterial activity in vitro. This may be due to retro-Michael reactions regenerating tetrodecamycin (1) as the biological active compound (Fig. 1).

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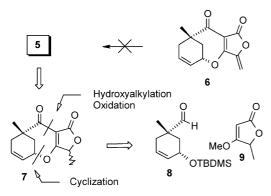
Figure 1.

We now report the synthesis of a tricyclic partial structure 5 of tetrodecamycin (1) comprising the unique anhydro  $\alpha$ -( $\gamma$ -hydroxy-acyl) tetronic acid derived core structure of the parent antibiotic.<sup>5</sup> The antibacterial activity of 5 as well as of a series of derivatives and analogues thereof was evaluated in vitro in an attempt to elucidate minimum structural requirements for antimicrobial potency. In addition, partial structures exhibiting an *exo*-methylene moiety were screened against human cell cultures in vitro in an effort to gauge the cytotoxicity of this class of compounds.

### **Results and Discussion**

#### Synthetic plan

Our synthetic strategy is outlined in Scheme 1. Tetrodecamycin partial structure 5 was envisioned to arise from tricyclic intermediate 7. This building block should be readily accessible from two key transformations: (a) regioselective 3-hydroxyalkylation of 4-*O*-methyl tetronate 9 with aldehyde 8 followed by an oxidation step to afford the respective 3-acyl-4-*O*-methyl tetronate<sup>6</sup> and (b) subsequent acid catalyzed cyclization of this precursor. In a previous communication we reported a short approach to tricyclic substructure 6, containing the crucial *exo*-methylene moiety.<sup>7</sup> In the course of further investigations, however, it became clear, that final



Scheme 1.

osmium tetroxide promoted *cis*-dihydroxylation of **6** to obtain partial structure **5** is unfeasible, due to an exclusive attack of the reagent on the *exo*-methylene group. Accordingly to obtain **5** from **7** it was obvious to introduce the *syn* diol group prior to the generation of the exocyclic C–C double bond.

## Synthesis of key building block 7

As depicted in Scheme 2, key building block 7 could be readily prepared in five steps from the known bicyclic lactone 10.8 First, 10 was treated with DIBAL in dichloromethane at -90 °C to give reduction product 11 in 86% yield. Selective silylation of the ring opened hydroxy aldehyde form 11b, which in dichloromethane solution prevails in an equilibrium with the corresponding bicyclic lactol 11a [11a/11b  $\sim$ 1:3, as determined by <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, rt)], was accomplished on treatment of 11 with TBDMSOTf in the presence of 2,6lutidine at low temperature (CH<sub>2</sub>Cl<sub>2</sub>, -78 °C). Thus, aldehyde 8 was obtained in high yield (94%) in addition to small amounts of the respective silvlated lactol. Cyclization precursor 14 (1:1 mixture of diastereomers) was obtained in 78% overall yield by highly regioselective 3-hydroxyalkylation of in situ generated boron furanolate 12,6 with aldehyde 8 and subsequent oxidation of the intermediate alcohol 13 (mixture of diastereomers) with 2-iodoxybenzoic acid (IBX).

To effect the desired cyclization, 3-acyl tetronate 14 was treated with catalytic amounts of concentrated H<sub>2</sub>SO<sub>4</sub> in dichloromethane at room temperature, thus affording

**Scheme 2.** (a) DIBAL, CH<sub>2</sub>Cl<sub>2</sub>, -90 °C (87%); (b) TBDMSOTf, lutidine, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C (94%); (c) (i) **9**, PhBCl<sub>2</sub>, *i*-Pr<sub>2</sub>NEt, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C; (ii) **8**, -78 °C to rt (86%); (d) IBX, DMSO, rt (91%); (e) cat concd H<sub>2</sub>SO<sub>4</sub>, CH<sub>2</sub>Cl<sub>2</sub>, rt (89%).

7 in high yield (89%). This acid-catalyzed ring closure step comprises the kinetically controlled initial formation of tetrahydrofuranylidene tetrahydrofuran-2,4-dione 15 (mixture of isomers), which slowly isomerizes to the thermodynamically favored product 7.9 The structure assigned for 7 was confirmed by IR and 2D NMR correlation with the respective 4-unsubstituted derivative, whose structure was unambiguously proven by X-ray diffraction analysis.<sup>5</sup>

# Synthesis of tetrodecamycin partial structure 5

Our initial efforts to dihydroxylate building block 7 were disappointing. Ruthenium-catalyzed *cis*-dihydroxylation according to Shing et al. <sup>10</sup> [RuCl<sub>3</sub> (0.07 equiv), NaIO<sub>4</sub> (1.5 equiv), MeCN, EtOAc, H<sub>2</sub>O, 0 °C] led to the desired diol **16** (as an inseparable 1:1 mixture of C-4 epimers) with good stereoselectivity (d.s = 95:5), but only in poor yield (27%), probably as a result of concurrent glycole cleavage (Scheme 3). On the other hand, commonly employed osmium tetroxide<sup>11</sup> or potassium permanganate<sup>12</sup> based dihydroxylations [e.g., OsO<sub>4</sub> (0.02 equiv), *N*-methylmorpholine *N*-oxide (1.0 equiv), acetone, *t*-BuOH, H<sub>2</sub>O, rt; KMnO<sub>4</sub>, 18-crown-6, CH<sub>2</sub>Cl<sub>2</sub>, rt) did not succeed at all, due to preferred oxidation of the acidic  $\gamma$ -methine group of the tetronate by these reagents.

To overcome this obstacle, it was obvious to block the acidic  $\gamma$ -position with an appropriate substituent, which both has to be compatible with standard *cis*-dihydroxylation conditions and should serve as an elimination precursor, suitable for the generation of the exocyclic C–C double bond.

We focused on the phenylsulfanyl group, since it is known to withstand osmium tetroxide under certain reaction conditions. <sup>13,14</sup> Moreover, this group serves as *syn* elimination precursors upon oxidation to the corresponding sulfoxide.

Introduction of the phenylsulfanyl group at the γ-position was accomplished by converting 7 first into the corresponding lithio furanolate 17 (LHMDS, THF, -78 °C) and then by reacting it with *N*-phenylmercaptophthalimide (-78 °C to rt) to give 18 (9:1 mixture of diastereomers) in 87% yield. Subsequent dihydroxylation with osmium tetroxide under stoichiometric conditions (pyridine, CCl<sub>4</sub>, rt) provided *cis*-diols 19 and 20 in high overall yield (87 and 6%, respectively) and with good diastereoselectivity (d.s. 19:20 = 93:7). The relative configurations of *cis*-diols 19 and 20 as well as that of 16 (vide supra) were determined on the basis of <sup>1</sup>H-<sup>1</sup>H *J*-coupling values and NOE studies. Next diol 19 was monoprotected as a *tert*-butyldimethylsilyl ether

Scheme 3. (a) RuCl<sub>3</sub> (0.07 equiv), NaIO<sub>4</sub> (1.5 equiv), MeCN, EtOAc, H<sub>2</sub>O, 0°C, (27%); (b) (i) LHMDS, THF, -78°C; (ii) *N*-phenyl-mercaptophthalimide, -78°C to rt (87%); (c) (i) OsO<sub>4</sub>, pyridine, CCl<sub>4</sub>, rt; (ii) aq NaHSO<sub>3</sub> (19: 87%; 20:%); (d) TBDMSOTf, lutidine, CH<sub>2</sub>Cl<sub>2</sub>, -78°C (92%); (e) 2-naphthoylchloride; NEt<sub>3</sub>, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, rt (22: 35%; 23: 18%; 24: 15%); (f) MCPBA, CH<sub>2</sub>Cl<sub>2</sub>, -20°C; (g) BaCO<sub>3</sub>, benzene, 80°C (25: 57% from 21); (26: 22% from 20); (27: 28% from 22); (28: 40% from 23); (29: 19% from 24); (6: 51% from 18); (h) HF, MeCN, 0°C (100%).

(TBDMSOTf, lutidine, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C) to form **21** regioselectively (92% yield). This additional step was necessary to enable isolation and purification of the subsequent products, which were hardly soluble in appropriate organic solvents when derived from **19**. Sulfide **21** was readily oxidized to the corresponding sulfoxide (mixture of diastereomers) by 3-chloroperoxybenzoic acid (CH<sub>2</sub>Cl<sub>2</sub>, -20 °C), which on pyrolysis (BaCO<sub>3</sub>, benzene, 80 °C) gave *exo*-methylene derivative **25** in 57% overall yield. Finally, deprotection of the hydroxyl group (HF, MeCN, 0 °C) afforded tetrodecamycin partial structure **5** in quantitative yield.

In the same way, diol **20** as well as 2-naphthoyl derivatives **22**, **23** and **24** were converted into the corresponding *exo*-methylene compounds **26**, **27**, **28** and **29** (in 22, 28, 40, and 19% overall yields, respectively). Esters **22**, **23** and **24** had been obtained in 35, 18 and 15% yield, respectively, from **19** by acylation with 2-naphthoyl chloride in the presence of catalytic amounts of DMAP (NEt<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, rt).

Finally, key building block **18** was converted into tetrodecamycin substructure **6** (51% yield). Indeed, this current approach was more convenient than our previously reported.<sup>7</sup>

#### Evaluation of in vitro antibacterial activities

The in vitro antibacterial activities of compounds 5–7, 16 and 26–29 were evaluated at concentrations from 0.5 to 128 μg mL<sup>-1</sup> against strains of two Gram-positive bacterial species: *S. aureus* [ATCC 25923, ATCC 29213 (MRSA)] and *E. faecalis* (ATCC 29212). The observed minimum inhibitory concentrations are listed in Table 1.

Tetrodecamycin partial structure **5**, which lacks a methyl group and in particular one cyclohexane ring of the *trans*-decalin fragment of the parent antibiotic, did not show growth inhibitory activity against the tested strains of *S. aureus* and *E. faecalis*, even at concentrations as high as 128 μg mL<sup>-1</sup>. This may be due to a loss of lipophilicity as compared with **1** or even more likely to a different geometry of the cyclohexane substructure included in both molecules. In case of **1** this ring, as part of the *trans*-decalin fragment, exists in the boat form, in case of partial structure **5**, however, it is chairlike. As a consequence the spatial orientation of the hydroxyl groups in both structures is also markedly different.

Table 1. In vitro antibacterial activity  $MIC^a$  (µg  $mL^{-1}$ )

Compd		S. aureus S. aureus <sup>b</sup> ATCC 25923 ATCC 29213			E. faecalis ATCC 29212	
5	>	128	>	128	>	128
6		4		8		4
7	>	128			>	128
16	>	128	>	128	>	128
26	>	128	>	128		64
27		8		16		4
28		128	>	128		64
29	>	128		_	>	128

<sup>&</sup>lt;sup>a</sup>Minimum inhibitory concentrations.

Interestingly a weak activity against the *E. faecalis* strain, but not against the *S. aureus* strains, could be noticed for diastereomer **26**. As was to be expected no activity was observed for dihydrotetrodecamycin partial structure **16**.

In case of tetrodecamycin (1) a clear enhancement of antimicrobial activity has been reported for certain 14-O substituents, particularly for a 2-naphthoyl residue (derivative 2, vide supra). Indeed, acylation of the respective hydroxyl group in 5 with 2-naphthalene carboxylic acid now led to weakly antibacterial active compound 28. A 16-fold higher activity, however, could be observed for 2-naphthoyl derivative 27, in which the neighbouring hydroxyl group has been acylated. This marked difference in activity may be due to the spatial orientation of the acyl residue, which in 27 is equatorial, just like for tetrodecamycin derivative 2, but is axial in case of compound 28 (vide supra). Notably, diacyl derivative 29 is bare of antimicrobial activity, indicating that a free, particularly axial aligned hydroxyl group may be relevant for efficacy.

Surprisingly we also found distinct antibacterial activity for tetrodecamycin substructure **6**, which lacks the vicinal *syn* diol function. Thus **6** had MICs of 4–8  $\mu$ g mL<sup>-1</sup> against *S. aureus* and *E. faecalis* suggesting a promising antibacterial potential. On the other hand dihydro derivative 7 was found to be inactive emphasizing the crucial role of the *exo*-methylene moiety for antibacterial activity.

### In vitro cytotoxicity of compounds 5-7 and 27

The cytotoxicity of tetrodecamycin partial structure 5 and of compounds 6, 7 and 27 was assessed in vitro against human leukemia HL-60 cells and human T cell leukemia Jurkat cells. The observed  $IC_{50}$  values are listed in Table 2.

Remarkably, the cytotoxicity of these compounds parallels with their antimicrobial activity. Thus for **5** at concentrations as high as  $100\,\mu\text{M}$  (26.5  $\mu\text{g}$  mL<sup>-1</sup>) no cytotoxicity was observed. On the other hand distinct activity against the two tested cell lines (IC<sub>50</sub> 10–23  $\mu\text{M}$ ) was found for compounds **6** and **27**. In contrast to **6** compound **7** proved to be inactive, evidently due to the absence of the *exo*-methylene moiety.

In conclusion, we have developed an efficient approach to the core structure 5 of the novel polyketide antibiotic

Table 2. Cytotoxicity of compounds 5–7 and 27

Compd	IC <sub>50</sub> (	μ <b>M</b> ) <sup>a</sup>	
	HL-60	Jurkat	
5	> 100	> 100	
6	10	23	
7	> 100	> 100	
27	N.D.	22	

<sup>&</sup>lt;sup>a</sup>IC<sub>50</sub>, 50% inhibitory concentration represents the mean from dose response curves of at least three experiments.

bMRSA.

tetrodecamycin (1). In addition to 5, diastereomer 26, analogue 6 and several derivatives (16, 27–29) have been prepared and evaluated for their antibacterial and cytotoxic activities. While tetrodecamycin partial structure 5 did not inhibit the growth of the tested Gram-positive pathogenes, 2-naphthoyl derivative 27 and analogue 6 showed promising antibacterial activities. However, these activities were paralleled by appreciable cytotoxicity. In summary the reactive *exo*-methylene moiety was shown to be crucial, but not sufficient by its own, for both antibacterial and cytotoxic activities. Further investigations, with the objective of determining whether the antimicrobial and cytotoxic activities could be detached from each other, are in progress in our laboratories.

#### **Experimental**

#### Chemistry

Unless otherwise noted, reactions were carried out in oven-dried glassware under an atmosphere of dry  $N_2$ . All reagents were used as commercially available. CH<sub>2</sub>Cl<sub>2</sub> and i-Pr<sub>2</sub>NEt were distilled from CaH<sub>2</sub> and THF from sodium metal immediately before use. Standard syringe techniques were applied for transferring anhydrous solvents. Flash chromatography: silica gel (Merck 60, 0.040-0.063 mm). Preparative HPLC: L-6000 pump, L-4000 UV/Vis, D-2000 Chromato Integrator (Merck-Hitachi), column, Hibar RT LiChrosorb<sup>®</sup> Si 60 (7  $\mu$ m, 250  $\times$  25 mm) (Merck). Melting points (mp, uncorrected values): Melting point apparatus according to Dr. Tottoli (Büchi no. 510). <sup>1</sup>H and <sup>13</sup>C NMR spectra: Eclipse 500 FTNMR spectrometer (Jeol), 500 and 125 MHz, respectively, chemical shifts  $(\delta)$  are reported in ppm, TMS as internal standard. IR spectra: FT-IR spectrometer Paragon 1000 (Perkin-Elmer). Mass spectra: 5989 Mass spectrometer with 59980 B particle beam LC/MS interface (Hewlett Packard). High resolution mass spectrometry: MStation 700 (Jeol). Elemental analysis: CHN Rapid (Heraeus).

(1RS,5RS)-1-Methyl-6-oxabicyclo[3.2.1]oct-3-ene-7-ol (1RS,5RS)-5-Hydroxy-1-methylcyclohex-3-enecarbaldehyde (11b). DIBAL (1.0 M in CH<sub>2</sub>Cl<sub>2</sub>, 22.0 mL, 22.0 mmol) was added dropwise to a solution of 10  $(2.763 \,\mathrm{g}, 20.0 \,\mathrm{mmol})$  in  $\mathrm{CH_2Cl_2}$   $(100 \,\mathrm{mL})$  at  $-90 \,\mathrm{^{\circ}C}$ . After stirring for 6 h at -90 °C the reaction was quenched by the addition of a saturated aqueous solution of potassium sodium tartrate (20 mL). The mixture was allowed to attain ambient temperature and was stirred at that temperature for 1.5 h. The layers were separated, and the aqueous layer was extracted with  $CH_2Cl_2$  (5 × 20 mL). The combined organic layers were dried (MgSO<sub>4</sub>) and evaporated under reduced pressure. The crude product was purified by flash chromatography  $(n-\text{hexane-Et}_2\text{O}, 4:6)$  to give 11 (2.430 g, 86.7%) as a colorless oil.

IR (film) 3390, 3030, 2927, 1728, 1716, 1652 cm  $^{-1}$ ; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 141 (3) [M+H<sup>+</sup>], 93 (100);  $^{1}$ H NMR (CDCl<sub>3</sub>) [In CDCl<sub>3</sub> the ring opened hydroxy aldehyde prevails in an equilibrium mixture with the

corresponding diastereomeric tricyclic lactols. Hydroxy aldehyde/lactol (major isomer)/lactol (minor isomer) = 75:17:8]:  $\delta$  = 1.18 (s, 0.75 × 3H, CH<sub>3</sub>), 1.20 (s,  $0.17 \times 3H$ , CH<sub>3</sub>), 1.22 (s, 0.08 × 3H, CH<sub>3</sub>), 1.66 (d, 0.17)  $\times$  1H, J = 10.8 Hz, 8-H), 1.77–1.87 (m, 0.08  $\times$  1H, 8-H), 1.80 (dd, 0.75  $\times$  1H, J = 5.4, 14.2 Hz, 6-H), 1.84 (dd,  $0.75 \times 1$ H, J = 2.1, 17.5 Hz, 2-H), 1.96–2.05 (m, 0.08 × 2H, 2-H and 8-H), 2.00 (dd,  $0.75 \times 1$ H, J = 3.5, 14.2 Hz, 6-H), 2.01 (dd, 0.17  $\times$  1H, J = 5.2, 10.8 Hz, 8-H), 2.18 (br d,  $0.17 \times 1$ H, J = 18.6 Hz, 2-H), 2.30 (dt,  $0.17 \times 1$ H, J=2.9, 18.6 Hz, 2-H), 2.44 (dd, 0.75 × 1H, J=2.7, 17.5 Hz, 2-H), 2.53 (d,  $0.08 \times 1$ H, J = 19.3 Hz, 2-H), 2.66 (br s,  $0.75 \times 1H$ , OH), 3.28 (br s,  $0.17 \times 1H$ , OH), 3.41 (br s,  $0.08 \times 1H$ , OH), 4.19 (br t,  $0.75 \times 1H$ , J = 5.4 Hz, 5-H), 4.49 (t, 0.17 × 1H, J = 5.2 Hz, 5-H),  $4.71 \text{ (m, } 0.08 \times 1\text{H, } 5\text{-H)}, 4.99 \text{ (s, } 0.17 \times 1\text{H, } 7\text{-H)}, 5.18$  $(s, 0.08 \times 1H, 7-H), 5.72 (m, 0.17 \times 1H, 3-H), 5.75-5.79$  $(m, 0.75 \times 2H, 3-H \text{ and } 4H), 5.80-5.90 (m, 0.08 \times 1H)$ 3-H), 6.16 (m,  $0.08 + 0.17 \times 1H$ , 4-H), 9.43 (s,  $0.75 \times 1H$ 1H, CHO). <sup>13</sup>C NMR (CDCl<sub>3</sub>) (only data for **11b** are shown):  $\delta = 21.33$  (CH<sub>3</sub>), 31.21 (C-2), 38.19 (C-6), 44.64 (C-1), 63.73 (C-5), 126.90 (C-4), 129.56 (C-3), 205.20 (CHO). Anal. calcd for C<sub>8</sub>H<sub>12</sub>O<sub>2</sub> (140.18): C, 68.55; H, 8.63. Found: C, 68.58; H, 8.65.

(1RS,5RS)-5-(tert-Butyldimethylsilanyloxy)-1-methylcyclohex-3-enecarbaldehyde (8). TBDMSOTf (3.44 mL, 14.98 mmol) was added dropwise to a solution of 11 (2.00 g, 14.27 mmol) and 2,6-lutidine (1.91 g, 2.07 mL, 17.83 mmol) in  $CH_2Cl_2$  (100 mL) at -78 °C. After stirring for 4 h at -78 °C the reaction mixture was poured into saturated aqueous sodium hydrogencarbonate (50 mL), and the resultant layers were separated. The aqueous layer was extracted with  $CH_2Cl_2$  (5 × 50 mL), and the combined organic layers were dried (MgSO<sub>4</sub>) and evaporated under reduced pressure. The crude product was purified by flash chromatography ( $CH_2Cl_2$ ) to give 8 (3.40 g, 93.6%) as a colorless oil.

IR (film) 3030, 2929, 1728 cm  $^{-1}$ ; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 255 (7) [M+H<sup>+</sup>], 123 (100);  $^{1}$ H NMR (CDCl<sub>3</sub>):  $\delta$  = 0.05 (s, 3H, SiCH<sub>3</sub>), 0.06 (s, 3H, SiCH<sub>3</sub>), 0.87 [s, 3H, CH<sub>3</sub>)<sub>3</sub>C], 1.06 (s, 3H, CH<sub>3</sub>), 1.77 (dd, 1H, J = 4.5, 13.5 Hz, 6-H), 1.78 (d, 1H, J = 18.2 Hz, 2-H), 1.95 (dd, 1H, J = 4.5, 13.5 Hz, 6-H), 2.52 (d, 1H, J = 18.2 Hz, 2-H), 4.23 (m, 1H, 5-H), 5.68 (m, 1H, 4-H), 5.77 (m, 1H, 3-H), 9.50 (s, 1H, CHO).  $^{13}$ C NMR (CDCl<sub>3</sub>):  $\delta$  = -4.8 (SiCH<sub>3</sub>), -4.5 (SiCH<sub>3</sub>), 18.1 [C(CH<sub>3</sub>)<sub>3</sub>], 21.4 (CH<sub>3</sub>), 26.9 [C(CH<sub>3</sub>)<sub>3</sub>], 30.7 (C-2), 39.7 (C-6), 44.3 (C-1), 64.4 (C-5), 127.0 (C-4), 129.2 (C-3), 204.0 (CHO). Anal. calcd for C<sub>14</sub>H<sub>26</sub>O<sub>2</sub>Si (254.45): C, 66.08; H, 10.30. Found: C, 65.94; H, 10.24.

(5RS)-3-[(1RS,5RS)-5-(tert-Butyldimethylsilanyloxy)-1-methylcyclohex-3-enecarbonyl]-4-methoxy-5-methyl-2(5H)-furanone and (5SR)-3-[(1RS,5RS)-5-(tert-butyldimethylsilanyloxy)-1-methylcyclohex-3-enecarbonyl]-4-methoxy-5-methyl-2(5H)-furanone (14). PhBCl<sub>2</sub> (2.49 g, 2.03 mL, 15.75 mmol) and then i-Pr<sub>2</sub>NEt (3.87 g, 5.18 mL, 30.0 mmol) were added dropwise to a solution of  $9^{15}$  (1.92 g, 15.0 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (60 mL) at -78 °C. After stirring for 1 h at -78 °C a solution of 8 (3.82 g, 15.0 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (15.0 mL) was added, and the

mixture was allowed to warm to room temperature and stirred for 1.5 h. Saturated aqueuos sodium hydrogencarbonate (30.0 mL) was added, and the mixture was extracted with  $CH_2Cl_2$  (5 × 100 mL). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated under reduced pressure. The residue was purified by flash chromatography (n-hexane–Et<sub>2</sub>O, 20:80) to give 13 (4.39 g, 85.8%) as an inseparable  $\sim 1:1:1:1$  mixture of diastereomers. A solution of 2-iodoxybenzoic acid (IBX) (5.41 g, 19.30 mmol) in DMSO (20 mL) was added to this mixture of diastereomers. The mixture was stirred at room temperature until TLC control indicates complete conversion ( $\sim 1.5 \, h$ ). The resulting suspension was diluted with Et<sub>2</sub>O (100 mL), applied on a silica gel column (~250 mL) and eluated with n-hexane-Et<sub>2</sub>O (1:1) to give **14** [4.45 g, 78% (from **8**)] as colorless crystals (inseparable 1:1 mixture of diastereomers).

IR (KBr) 3041, 2952, 2929, 2858, 1750, 1681, 1629 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 381 (1) [M+H<sup>+</sup>], 249 (100); <sup>1</sup>H NMR (CDCl<sub>3</sub>): diastereomer 1:  $\delta = 0.08$  (s, 3H, CH<sub>3</sub>Si), 0.09 (s, 3H, SiCH<sub>3</sub>), 0.89 [s, 3H, (CH<sub>3</sub>)<sub>3</sub>C], 1.38 [s, 3H,  $CH_3$ -(C-1')], 1.49 [d, J = 6.8 Hz, 3H,  $CH_3$ -(C-5)], 1.83 (dd, J = 12.6/8.7 Hz, 1H, 6'-H), 2.09–2.22 (m, 2H, 6'-H and 2'-H), 2.45-2.55 (m, 1H, 2'-H), 3.85 (s, 3H, OCH<sub>3</sub>), 4.35 (m, 1H, 5'-H), 4.82 (q, J = 6.8 Hz, 1H, 5-H), 5.63 (m, 1H, 4'-H), 5.68 (m, 1H, 3'-H); diastereomer 2:  $\delta = 0.09$  (s, 3H, CH<sub>3</sub>Si), 0.10 (s, 3H, SiCH<sub>3</sub>), 0.90 [s, 3H, (CH<sub>3</sub>)<sub>3</sub>C], 1.37 [s, 3H, CH<sub>3</sub>-(C-1')], 1.49 [d, J = 6.8 Hz, 3H, CH<sub>3</sub>-(C-5)], 1.71 (dd, J = 12.6/8.7 Hz, 1H, 6'-H), 2.09–2.22 (m, 2H, 6'-H and 2'-H), 2.45–2.55 (m, 1H, 2'-H), 3.85 (s, 3H, OCH<sub>3</sub>), 4.35 (m, 1H, 5'-H), 4.80 (q,  $J = 6.8 \,\mathrm{Hz}$ , 1H, 5-H), 5.63 (m, 1H, 4'-H), 5.68 (m, 1H, 3'-H). Anal. calcd for  $C_{20}H_{32}O_5Si$  (380.56): C, 63.12; H, 8.48. Found: C, 63.25; H, 8.65.

(1RS,4RS,9RS)-4,9-Dimethyl-2,5-dioxa-tricyclo[7.3.1.0<sup>3,7</sup>]-trideca-3(7),11-diene-6,8-dione and (1RS,4SR,9RS)-4,9-dimethyl-2,5-dioxa-tricyclo[7.3.1.0<sup>3,7</sup>]trideca-3(7),11-diene-6,8-dione (7). Sulphuric acid (96%, 1.64 g, 0.90 mL, 16.75 mmol) was added to a solution of 14 (2.55 g, 6.70 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (250 mL) at room temperature. After stirring for 1.5 h the reaction mixture was poured into H<sub>2</sub>O (60 mL). The layers were separated, and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (5 × 150 mL). The combined organic layers were washed with H<sub>2</sub>O (2 × 50 mL), dried (MgSO<sub>4</sub>) and evaporated under reduced pressure. The crude product was purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>-EtOAc, 80:20) to give 7 (1.398 g, 89.1%) as colorless crystals (inseparable 1:1 mixture of diastereomers).

IR (KBr) 3037, 2979, 2932, 1762, 1650, 1599 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub>+): m/z (%) = 235 (100) [M+H+]; <sup>1</sup>H NMR (CDCl<sub>3</sub>): diastereomer 1:  $\delta$ =1.21 [s, 3H, CH<sub>3</sub>-(C-9)], 1.44 [d, J=6.7 Hz, 3H, CH<sub>3</sub>-(C-4)], 1.96 (d, J=17.9 Hz, 1H, 10-H), 2.10 (dd, J=16.4/6.4 Hz, 1H, 13-H), 2.46 (d, J=16.4 Hz, 1H, 13-H), 2.73 (d, J=17.9 Hz, 1H, 10-H), 4.80 (q, J=6.7 Hz, 1H, 4-H), 5.29 (m, 1H, 1-H), 5.71 (m, 1H, 12-H), 6.17 (m, 1H, 11-H); diastereomer 2:  $\delta$ =1.22 [s, 3H, CH<sub>3</sub>-(C-9)], 1.53 [d, J=6.8 Hz, 3H, CH<sub>3</sub>-(C-4)], 1.96 (d, J=17.9 Hz, 1H, 10-H), 2.10 (dd, J=16.4/6.4 Hz, 1H, 13-H), 2.46 (d, J=16.4 Hz, 1H,

13-H), 2.73 (d, J = 17.9 Hz, 1H, 10-H), 4.69 (q, J = 6.8 Hz, 1H, 4-H), 5.30 (m, 1H, 1-H), 5.72 (m, 1H, 12-H), 6.17 (m, 1H, 11-H). Anal. calcd for  $C_{13}H_{14}O_4$  (234.25): C, 66.66; H, 6.02. Found: C, 66.68; H, 5.93.

(1RS,4RS,9RS,11SR,12SR)-11,12-Dihydroxy-4,9-dimethyl-2,5-dioxatricyclo[7.3.1.O<sup>3,7</sup>]trideca-3(7)-ene-6,8-dione and (1RS,4SR,9RS,11SR,12SR)-11,12-dihydroxy-4,9-dimethyl-2,5-dioxatricyclo[7.3.1.O <sup>3,7</sup>]trideca-3(7)-ene-6,8dione (16). Ruthenium trichloride (0.6 mg, 2.3 µmol) and a solution of sodium periodate (10.7 mg,  $0.05 \,\mathrm{mmol}$ ) in H<sub>2</sub>O (66  $\mu$ L) were added to a solution of 7 (7.8 mg, 0.033 mmol) in EtOAc-CH<sub>3</sub>CN, 1:1 (0.4 mL) at 0°C. The resulting brown solution was stirred at 0°C for 3 min. Then the reaction was quenched by the addition of a 10% aqueous solution of sodium bisulfite (1.0 mL). The mixture was extracted with EtOAc (5  $\times$ 5 mL), and the combined organic layers were dried (MgSO<sub>4</sub>) and evaporated under reduced pressure to obtain the crude product as a complex mixture of diastereomers (95:5 with respect to C11-C12 and 1:1 with respect C-4 in each case as determined by <sup>1</sup>H NMR). The crude product was purified by flash chromatography (nhexane-EtOAc, 60:40) to obtain 16 (2.4 mg, 27.0%) as colorless crystals (1:1 mixture of C-4 epimers).

IR (KBr) 3491, 2923, 2847, 1754, 1648, 1603 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>) m/z (%) = 269 (100) [M+H<sup>+</sup>]; <sup>1</sup>H NMR (Acetone- $d_6$ ):  $\delta$  = 1.13 (s, 3H, 9-CH<sub>3</sub>), 1.44 (d, 0.5 × 3H, J = 6.8 Hz, 4-CH<sub>3</sub>), 1.45 (d, 0.5 × 3H, J = 6.8 Hz, 4-CH<sub>3</sub>), 1.74–1.79 (m, 2H, 10-H), 2.12 (dd, 1H, J = 4.6, 16.4 Hz, 13-H), 2.33 (d, 0.5 × 1H, J = 16.4 Hz, 13-H), 2.35 (d, 0.5 × 1H J = 16.4 Hz, 13-H), 3.59 (m, 1H, 11-H); <sup>13</sup>C NMR (acetone- $d_6$ ):  $\delta$  = 17.4, 17.6 (4-CH<sub>3</sub>), 26.3, 26.4 (9-CH<sub>3</sub>), 37.2, 37.3 (C-13), 39.8, 40.0 (C-10), 47.8, 47.9 (C-9), 65.75, 65.84 (C-11), 70.2, 70.3 (C-12), 72.6, 72.7 (C-5), 84.23, 84.24 (C-1); HRMS (EI): m/z calcd for C<sub>13</sub>H<sub>16</sub>O<sub>6</sub>: 268.0947; Found: 268.0990.

(1RS,4RS,9RS)-4,9-Dimethyl-4-phenylsulfanyl-2,5-dioxatricyclo7.3.1.O<sup>3,7</sup>trideca-3(7),11-dien-6,8-dione (18). A 1.0 M solution of LiHMDS in THF (6.14 mL, 6.14 mmol) was added dropwise to a suspension of 7  $(1.370 \,\mathrm{g}, 5.85 \,\mathrm{mmol})$  in THF  $(25 \,\mathrm{mL})$  at  $-78 \,^{\circ}\mathrm{C}$ . The resulting orange solution was stirred at -78 °C for 1.5 h before a solution of N-phenylmercaptophthalimide (1.57 g, 6.14 mmol) in THF (25 mL) was added dropwise. After stirring at -78 °C for 1 h the reaction mixture was allowed to attain room temperature before it was poured into potassium dihydrogenphosphate buffer pH 4.5 (50 mL). The mixture was extracted with Et<sub>2</sub>O (5 × 100 mL). The combined organic layers were washed with saturated aqueous sodium hydrogenicarbonate (2  $\times$ 50 mL), dried (MgSO<sub>4</sub>) and evaporated under reduced pressure. The crude product (90:10 mixture of diastereomers as determined by <sup>1</sup>H NMR) was purified by flash chromatography (n-hexane-EtOAc, 60:40) and prep. HPLC (n-heptane-EtOAc, 60:40, 15 mL/min, 275 nm,  $R_t = 23.9 \,\text{min}$ ) to give **18** (1.735 g, 86.7%) as colorless crystals; mp 176 °C.

IR (KBr) 3051, 2960, 2927, 1773, 1655, 1603 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 343 (100) [M+H<sup>+</sup>]; <sup>1</sup>H NMR

(CDCl<sub>3</sub>):  $\delta$  = 0.88 [s, 3H, CH<sub>3</sub>-(C-9)], 1.72–1.88 (m, 3H, 10-H and 13-H), 1.78 [s, 3H, CH<sub>3</sub>-(C-4)], 2.59 (dd, 1H, J= 17.8/6.2 Hz, 10-H), 5.29 (m, 1H, 1-H), 5.67 (m, 1H, 12-H), 6.10 (m, 1H, 11-H), 7.31–7.41 (m, 3H, H<sub>arom</sub>), 7.56–7.60 (m, 2H, H<sub>arom</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  = 23.5 [CH<sub>3</sub>-(C-4)], 25.4 [CH<sub>3</sub>-(C-9)], 34.1 (C-13), 37.2 (C-10), 44.6 (C-9), 76.4 (C-1), 88.6 (C-4), 102.9 (C-7), 121.4 (C-12), 128.4, 129.2, 130.3, 136.4 (C<sub>arom</sub>), 135.5 (C-11), 165.4 (C-6), 177.1 (C-3), 198.0 (C-8). Anal. calcd for C<sub>19</sub>H<sub>18</sub>O<sub>4</sub>S (342.42): C, 66.65; H, 5.30; S, 9.36. Found: C, 66.66; H, 5.23; S, 9.30.

(1RS,4RS,9RS,11SR,12SR)-11,12-Dihydroxy-4,9-dimethyl-4-phenylsulfanyl-2,5-dioxa tricyclo[7.3.1.0 <sup>3,7</sup>]tridec-3(7)-en-6,8-dione (19) and (1RS,4RS,9RS,11RS,12RS)-11,12-dihydroxy-4,9-dimethyl-4-phenylsulfanyl-2,5-dioxa tricyclo[7.3.1.O <sup>3,7</sup>]tridec-3(7)-en-6,8-dione (20). A solution of osmium tetroxide (1.134 g, 4.46 mmol) in CCl<sub>4</sub> (30 mL) was added to a solution of 18 (1.45 g, 4.25 mmol) in pyridine (90 mL) at room temperature. The resulting brown solution was stirred for 2 h before a 10% aqueous solution of sodium sulfite (100 mL) and pyridine (30 mL) were added. After stirring for 1 h the layers were separated, and the aqueous layer was extracted with  $CH_2Cl_2$  (5 × 120 mL). The combined organic layers were dried (MgSO<sub>4</sub>) and evaporated under reduced pressure to give a crude product as a mixture of diastereomers (19:20 = 93.7 as determined by <sup>1</sup>H NMR). The crude product was purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>–EtOAc–2-propanol, 30:70:1) to give **19** (1.394 g, 87.1%) and **20** (91 mg, 5.7%) as colorless crystals; mp 149 and 229 °C, respectively.

19. IR (KBr) 3439, 2932, 1766, 1659, 1607 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 377 (66) [M+H<sup>+</sup>], 269 (100); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$ =0.96 [s, 3H, CH<sub>3</sub>-(C-9)], 1.39 (d, J=16.6 Hz, 1H, 13-H), 1.66 (dd, J=12.7/12.6 Hz, 1H, 10-H<sub>ax</sub>), 1.79 (dd, J=12.7/4.0 Hz, 1H, 10-H<sub>eq</sub>), 1.84 [s, 3H, CH<sub>3</sub>-(C-4)], 1.90 (dd, J=16.6/4.6 Hz, 1H, 13-H), 2.71 (d, J=4.5 Hz, 1H, OH), 2.80 (d, J=2.0 Hz, 1H, OH), 3.58 (m, 1H, 11-H), 4.13 (m, 1H, 12-H), 5.09 (m, 1H, 1-H), 7.30–7.42 (m, 3H, H<sub>arom</sub>), 7.50–7.54 (m, 2H, H<sub>arom</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$ =23.8 [CH<sub>3</sub>-(C-4)], 26.4 [CH<sub>3</sub>(C-9)], 29.2 (C-13), 39.3 (C-10), 47.6 (C-9), 66.1 (C-11), 70.4 (C-12), 83.3 (C-1), 89.5 (C-4), 102.5 (C-7), 128.3, 129.5, 130.7, 136.4 (C<sub>arom</sub>), 165.6 (C-6), 179.2 (C-3), 197.9 (C-8); HRMS: Anal. calcd for C<sub>19</sub>H<sub>20</sub>O<sub>6</sub> (M<sup>+</sup>): 376.0981; Found: 376.1114.

**20**. IR (KBr) 3461, 3302, 2927, 1754, 1648, 1616 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 377 (28) [M+H<sup>+</sup>], 111 (100); <sup>1</sup>H NMR (acetone- $d_6$ ):  $\delta$  = 0.82 [s, 3H, CH<sub>3</sub>-(C-9)], 1.60 (dd, J=14.4/2.3 Hz, 1H, 10-H), 1.64 (dt, J=16.5/2.9 Hz, 1H, 13-H), 1.75 [s, 3H, CH<sub>3</sub>-(C-4)], 1.84 (dd, J=16.5/3.8 Hz, 1H, 13-H), 2.18 (dt, J=14.4/3.3 Hz, 1H, 10-H), 3.76 (dt, J=8.8/3.9 Hz, 1H, 12-H), 3.92 (ddd, J=3.9/3.3/2.3 Hz, 1H, 11-H), 3.99 [d, J=2.9 Hz, 1H, OH-(C-12)], 4.02 [d, J=8.8 Hz, 1H, OH-(C-11)], 5.14 (dd, J=3.9/2.3 Hz, 1H, 4-H), 7.41 (m, 2H<sub>arom</sub>), 7.46 (m, 1H, H<sub>arom</sub>), 7.57 (m, 2H, H<sub>arom</sub>). <sup>13</sup>C NMR (acetone- $d_6$ ):  $\delta$ =23.8 [(C-4)-CH<sub>3</sub>], 26.6 [(C-9)-CH<sub>3</sub>], 33.6 (C-13),

43.0 (C-10), 44.8 (C-9), 68.4 (C-11), 71.4 (C-12), 83.1 (C-1), 89.6 (C-4), 102.8 (C-7), 129.3, 129.7, 130.6, 136.7 (C<sub>arom</sub>), 164.8 (C-6), 177.9 (C-3), 198.0 (C-8); HRMS: Anal. calcd for  $C_{19}H_{20}O_6$  (M $^+$ ): 376.0981; Found: 376.1006.

(1RS,4RS,9RS,11SR,12SR)-11-(tert-Butyldimethylsilanyloxy)-12-hydroxy-4,9-dimethyl-4-phenylsulfanyl-2,5-dioxa-tricyclo[7.3.1.0<sup>3,7</sup>]tridec-3(7)-ene-6,8-dione (21). tert-Butyldimethylsilyl trifluoromethanesulphonate (775 mg, 630 µL, 2.766 mmol) was added dropwise to a solution of **19** (496 mg, 1.317 mmol) and 2,6-lutidine (353 mg,  $382 \,\mu\text{L}$ ,  $3.293 \,\text{mmol}$ ) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL) at  $-78 \,^{\circ}\text{C}$ . After stirring for 4 h at -78 °C the reaction mixture was poured into saturated aqueous sodium hydrogencarbonate (30 mL), and the resultant layers were separated. The aqueous layer was extracted with  $CH_2Cl_2$  (5 × 50 mL), the combined organic layers were dried (MgSO<sub>4</sub>) and evaporated under reduced pressure. The crude product was purified by flash chromatography (n-hexane–Et<sub>2</sub>O, 40:60) to give **21** (591 mg, 91.8%) as colorless crystals; mp 155 °C.

IR (KBr) 3485, 3061, 2952, 2931, 2855, 1792, 1767, 1669, 1607 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 491 (100)  $[M + H^{+}]$ ; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = -0.01$  (s, 3H, CH<sub>3</sub>Si), 0.00 (s, 3H, CH<sub>3</sub>Si), 0.84 [s, 9H, (CH<sub>3</sub>)<sub>3</sub>C], 0.98 [s, 3H,  $CH_3$ -(C-9)], 1.40 (d, J = 16.5 Hz, 1H, 13-H), 1.62 (dd, J = 13.1/12.3 Hz, 1H, 10-H<sub>ax</sub>), 1.80 (dd, J = 13.1/4.5 Hz, 1H, 10-H<sub>eq</sub>), 1.81 [s, 3H, CH<sub>3</sub>-(C-4)], 1.88 (dd, J = 16.5/ 4.6 Hz, 1H, 13-H), 2.56 (s<sub>br</sub>, 1H, OH), 3.50 (ddd, J = 12.3/4.5/3.0 Hz, 1H, 11-H), 3.94 (m, 1H, 12-H), 5.09 (m, 1H, 1-H), 7.31–7.43 (m, 3H, H<sub>arom</sub>), 7.51–7.57 (m, 2H,  $H_{arom}$ ); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = -5.1$  (CH<sub>3</sub>Si), 4.6  $(CH_3Si)$ , 18.8  $[(CH_3)_3C]$ , 23.8  $[CH_3-(C-4)]$ , 25.6  $[(CH_3)_3C]$ , 26.4  $[CH_3-(C-9)]$ , 29.0 (C-13), 39.3 (C-10), 67.1 (C-11), 71.0 (C-12), 82.1 (C-1), 89.1 (C-4), 102.7 (C-7), 128.1, 129.3, 130.5, 136.2 (C<sub>arom</sub>), 154.9 (C-6), 178.7 (C-3), 197.2 (C-9). Anal. calcd for C<sub>25</sub>H<sub>34</sub>O<sub>6</sub>SSi (490.70): C, 61.19; H, 6.98; S, 6.53; Found: C, 61.24; H, 7.08; S, 6.84.

2-Naphthalene-2-carboxylic acid (1RS,4RS,9RS,11SR,12-SR)-12-hydroxy-4,9-dimethyl-6,8-dioxo-4-phenylsulfanyl-2,5-dioxatricyclo-[7.3.1.0<sup>3,7</sup>tridec-3(7)-en-11-yl ester (22) and 2-naphthalene-2-carboxylic acid (1RS,4RS,9RS,11-SR,12SR)-11-hydroxy-4,9-dimethyl-6,8-dioxo-4-phenylsulfanyl-2,5-dioxatricyclo-[7.3.1.0<sup>3,7</sup>tridec-3(7)-en-12-yl ester (23) and 2-naphthalene-2-carboxylic acid (1RS,9-RS,11SR,12SR)-4,9-dimethyl-12-(naphthalene-2-carbonyloxy)-6,8-dioxo-4-phenylsulfanyl-2,5-dioxatricyclo-[7.3.1.0<sup>3,7</sup>tridec-3(7)-en-11-yl ester (24). A solution of 2-naphthoyl chloride (76 mg, 0.4 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1.5 mL) was added dropwise to a solution of 19 (125 mg, 0.333 mmol) and N-ethyldiisopropylamine (56 mg, 75 μL, 0.433 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (3 mL) at room temperature during 4h. After stirring at room temperature for 12h saturated aqueous sodium hydrogencarbonate (3 mL) was added, and the resultant layers were separated. The aqueous layer was extracted with Et<sub>2</sub>O (5  $\times$ 15 mL). The combined organic layers were dried (MgSO<sub>4</sub>) and evaporated under reduced pressure. The crude product was purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>–EtOAc, 95:5) to give **22** (61.9 mg, 35.1%), **23** (31.6 mg, 17.9%) and **24** (35.1 mg, 15.4%) as colorless crystals; mp 258, 220–222 and 89  $^{\circ}$ C, respectively.

**22**. IR (KBr) 3480, 3055, 2934, 1763, 1721, 1603 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 1.10$  (s, 3H, 9-CH<sub>3</sub>), 1.54 (m, 1H, 10-H), 1.94 (s, 3H, 4-CH<sub>3</sub>), 2.04–2.13 (m, 3H, 10-H and 13-H), 2.34 (br. s, 1H, OH), 4.46 (m, 1H, 12-H), 5.00 (ddd, 1H, J = 2.6, 5.6, 11.8 Hz, 1-H), 5.15 (m, 1H, 11-H), 7.36 (m, 2H, H<sub>arom</sub>), 7.41 (m, 1H, H<sub>arom</sub>), 7.54–7.58 (m, 3H,  $H_{arom}$ ), 7.62 (m, 1H,  $H_{arom}$ ), 7.88 (m, 2H,  $H_{arom}$ ), 7.94 (br. d, 1H, J = 8.4 Hz,  $H_{arom}$ ), 7.96 (dd, 1H, J = 8.4 Hz,  $H_{arom}$ ), 7.96 (dd, 1H,  $H_{arom}$ J=1.7, 8.7 Hz, H<sub>arom</sub>), 8.53 (br. s, 1H, H<sub>arom</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 23.8 (4-CH<sub>3</sub>), 26.4 (9-CH<sub>3</sub>), 29.0 (C-10), 35.4 (C-13), 47.5 (C-9), 68.6 (C-12), 69.3 (C-1), 82.8 (C-11), 89.3 (C-4), 102.9 (C-7), 124.9, 126.4, 127.0, 127.9, 128.3, 128.4, 128.8, 129.0, 129.4, 130.5, 131.4, 131.4, 132.5, 135.9, 136.3, 164.8 (C<sub>arom</sub>), 165.4 (C-6), 178.7 (C-3), 196.2 (C-8); MS (CI, CH<sub>5</sub><sup>+</sup>): m/z = 531 (100)  $[M+H^+]$ ; HRMS (EI): m/z calcd for  $C_{30}H_{26}O_7S-$ C<sub>6</sub>H<sub>5</sub>S: 421.1287; Found: 421.1287.

**23**. IR (KBr) 3487, 3054, 2925, 1775, 1717, 1615 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 1.08$  (s, 3H, 8-CH<sub>3</sub>), 1.53 (d, 1H, J = 16.8 Hz, 13-H), 1.80 (dd, 1H, J = 11.3, 13.5 Hz, 10-H), 1.88 (s, 3H, 4-CH<sub>3</sub>), 1.95 (br. s, 1H, OH), 1.97 (dd, 1H, J=5.0, 16.8 Hz, 13-H), 2.24 (dd, 1H, J=5.0, 13.5 Hz, 10-H), 3.84 (m, 1H, 11-H), 5.24 (m, 1H, 1-H), 5.56 (m, 1H, 12-H), 7.31-7.36 (m, 2H, H<sub>arom</sub>), 7.39 (m, 1H, H<sub>arom</sub>), 7.52–7.56 (m, 2H, H<sub>arom</sub>), 7.60 (t, 1H,  $J = 7.6 \text{ Hz}, \text{ H}_{arom}$ ), 7.65 (t, 1H,  $J = 7.6 \text{ Hz}, \text{ H}_{arom}$ ), 7.92 (d, 1H,  $J=8.2 \,\text{Hz}$ ,  $H_{\text{arom}}$ ), 7.93 (d, 1H,  $J=8.6 \,\text{Hz}$ ,  $H_{arom}$ ), 7.99 (d, 1H,  $J=8.2 \, Hz$ ,  $H_{arom}$ ), 8.01 (dd, 1H, J=1.7, 8.6 Hz, H<sub>arom</sub>), 8.59 (br. s, 1H, H<sub>arom</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 24.0$  (4-CH<sub>3</sub>), 26.9 (9-CH<sub>3</sub>), 30.6 (C-13), 40.9 (C-10), 47.4 (C-9), 65.6 (C-11), 71.7 (C-12), 81.3 (C-1), 89.6 (C-4), 103.6 (C-7), 125.1, 126.1, 126.1, 127.2, 128.0, 128.4, 128.4, 128.8, 129.0, 129.5, 129.5, 130.5, 131.7, 132.5, 136.0, 136.3, 165.0 (C<sub>arom</sub>), 166.0 (C-6), 178.5 (C-3), 197.3 (C-8); MS (CI,  $CH_5^+$ ); m/z $(\%) = 531 (9) [M + H^{+}], 249 (100); HRMS (EI): m/z$ calcd for C<sub>30</sub>H<sub>26</sub>O<sub>7</sub>S-C<sub>6</sub>H<sub>5</sub>S: 421.1287; Found: 421.1287.

**24**. IR (KBr) 3061, 2925, 1784, 1720, 1610 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 1.19$  (s, 3H, 9-CH<sub>3</sub>), 1.71 (d, 1H,  $J = 16.7 \,\mathrm{Hz}$ , 13-H), 2.00 (s, 3H, 4-CH<sub>3</sub>), 2.14 (dd, 1H, J = 4.8, 16.7 Hz, 13-H), 2.18 (dd, 1H, J = 12.1, 13.2 Hz, 10-H), 2.34 (dd, 1H, J = 4.6, 13.2, 10-H), 5.23 (ddd, 1H, J = 3.0, 4.6, 12.1 Hz, 11-H), 5.34 (m, 1H, 1-H), 5.89 (m, 1H, 12-H), 7.34-7.40 (m, 2H, H<sub>arom</sub>), 7.41 (m, 1H, H<sub>arom</sub>), 7.45 (m, 1H, H<sub>arom</sub>), 7.54 (m, 1H, H<sub>arom</sub>), 7.56– 7.60 (m, 2H, H<sub>arom</sub>), 7.59 (m, 1H, H<sub>arom</sub>), 7.66 (m, 1H,  $H_{arom}$ ), 7.70 (d, 1H, J = 8.2 Hz,  $H_{arom}$ ), 7.77 (d, 1H,  $J = 8.7 \text{ Hz}, \text{ H}_{arom}$ ), 7.81 (d, 1H,  $J = 8.2 \text{ Hz}, \text{ H}_{arom}$ ), 7.86 (dd, 1H, J=1.7, 8.7 Hz,  $H_{arom}$ ), 7.91–7.98 (m, 3H,  $H_{arom}$ ), 8.01 (dd, 1H, J = 1.7, 8.6 Hz,  $H_{arom}$ ), 8.39 (br. s, 1H, H<sub>arom</sub>), 8.58 (br. s, 1H, H<sub>arom</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 23.8 \text{ (4-CH<sub>3</sub>)}, 26.7 \text{ (9-CH<sub>3</sub>)}, 30.3 \text{ (C-10)}, 37.2 \text{ (C-13)},$ 47.3 (C-9), 66.9 (C-11), 69.0 (C-12), 80.7 (C-1), 89.5 (C-

4), 103.2 (C-7), 124.9, 125.0, 126.1, 126.5, 126.8, 127.2, 127.8, 128.0, 128.4, 128.4, 128.6, 128.8, 129.0, 129.4, 129.5, 129.5, 130.6, 131.5, 131.8, 132.4, 132.5, 135.7, 136.0, 136.4, 164.7, 165.1 ( $C_{arom}$ ), 165.4 (C-6), 178.5 (C-3), 196.0 (C-8); MS (CI, CH<sub>5</sub>+): m/z (%) = 685 (100) [M+H+]; HRMS (EI): m/z calcd for  $C_{41}H_{32}O_8S-C_6H_5S$ : 575.1735; Found: 575.1706.

# Oxidative elimination of the phenylsulfanyl group—general procedure

A 0.1 M solution of 3-chloroperoxybenzoic acid in CH<sub>2</sub>Cl<sub>2</sub> (2.50 mL, 0.25 mmol) was added dropwise to a solution of the respective sulfide (0.25 mmol) in CH<sub>2</sub>Cl<sub>2</sub>  $(15 \,\mathrm{mL})$  at  $-20\,^{\circ}\mathrm{C}$ . After stirring for 1–4 h at  $-20\,^{\circ}\mathrm{C}$ (until TLC control indicates complete conversion) the reaction mixture was poured into saturated aqueous sodium hydrogenearbonate (25 mL), and the resultant layers were separated. The aqueous layer was extracted with Et<sub>2</sub>O or CH<sub>2</sub>Cl<sub>2</sub> (5  $\times$  50 mL). The combined organic layers were dried (MgSO<sub>4</sub>) and evaporated under reduced pressure. A suspension of the resulting crude product and barium carbonate (245 mg, 1.25 mmol) in benzene (50 mL) was refluxed for 30 min. The suspension was filtered, and the filtrate was evaporated under reduced pressure. The crude product was purified by flash chromatography.

(1*RS*,9*RS*,11*RS*,12*RS*) - 11 - (*tert* - Butyldimethyl - silanyloxy)-12-hydroxy-9-methyl-4-methylene-2,5-dioxatricyclo[7.3.1.0<sup>3,7</sup>]tridec-3(7)-ene-6,8-dione (25). Prepared according to general procedure from 21 (24.5 mg, 0.05 mmol). Purified by flash chromatography (*n*-hexane–EtOAc, 70:30) to give 25 (11 mg, 57.7%) as colorless crystals; mp 155–158 °C.

IR (KBr) 3487, 2928, 2856, 1794, 1673, 1586 cm<sup>-1</sup>; MS (CI,CH<sub>5</sub><sup>+</sup>): m/z (%) = 381 (71) [M+H<sup>+</sup>], 257 (100);  $^{1}$ H NMR (CDCl<sub>3</sub>):  $\delta$ =0.03 (s, 3H, CH<sub>3</sub>Si), 0.04 (s, 3H, CH<sub>3</sub>Si), 0.86 [s, 9H, (CH<sub>3</sub>)<sub>3</sub>C], 1.25 [s, 3H, CH<sub>3</sub>-(C-9)], 1.75 (dd, J=13.0/12.2 Hz, 1H, 10-H<sub>ax</sub>), 1.93 (dd, J=13.0/4.5 Hz, 1H, 10-H<sub>eq</sub>), 2.17 (dd, J=16.7/4.5 Hz, 1H, 13-H), 2.25 (d, J=16.7 Hz, 1H, 13-H), 2.65 (s<sub>br</sub>, 1H, OH), 3.64 (ddd, J=12.2/4.5/3.1 Hz, 11-H), 4.01 (m, 1H, 12-H), 5.12 (m, 1H, 1-H), 5.30 [d, J=2.8 Hz, 1H, (C-4)=CH<sub>2</sub>], 5.38 [d, J=2.8 Hz, 1H, (C-4)=CH<sub>2</sub>]. Anal. calcd for: C<sub>19</sub>H<sub>28</sub>O<sub>6</sub>Si (380.52): C, 59.87; H, 7.42; Found: C, 60.14; H, 7.41.

(1RS,9RS,11SR,12SR)-11,12-Dihydroxy-9-methyl-4-methylene-2,5-dioxatricyclo-[7.3.1.0<sup>3,7</sup>tridec-3(7)-ene-6,8-dione (5). A 0.1 M HF solution in acetonitrile (660  $\mu$ L, 0.066 mmol) was added to a solution of 25 (8.4 mg, 0.022 mmol) in acetonitrile (1 mL) at 0 °C. After stirring at 0 °C for 21 h, a colorless precipitate has been formed, which was filtered off and was washed with pentane to give 5 (5.9 mg, 100%) as colorless crystals; mp > 370 °C.

IR (KBr) 3460, 2924, 1786, 1588 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 267 (100) [M+H<sup>+</sup>]; <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$ =1.07 [s, 3H, CH<sub>3</sub>-(C-9)], 1.58 (dd, J=12.4/3.7 Hz, 1H, 10-H<sub>eq</sub>), 1.72 (t, J=12.4 Hz, 1H, 10-H<sub>ax</sub>), 1.99 (dd,

J=16.4/4.5 Hz, 1H, 13-H), 2.31 (d, J=16.4 Hz, 1H, 13-H), 3.39 (d, J=12.4 Hz, 1H, 11-H), 3.89 (m, 1H, 12-H), 4.80 (d, J=5.6 Hz, 1H, OH), 4.98 (m, 1H, 1-H), 5.39 [d, J=2.8 Hz, 1H, (C-4)=CH<sub>2</sub>], 5.41 [d, J=2.8 Hz, 1H, (C-4)=CH<sub>2</sub>], 5.43 (d, J=4.5 Hz, 1H, OH); <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta=26.3$  [CH<sub>3</sub>-(C-9)], 28.4 (C-13), 39.2 (C-10), 47.7 (C-9), 64.7 (C-11), 72.3 (C-12), 84.2 (C-1), 96.7 [(C-4)=CH<sub>2</sub>], 101.5 (C-7), 147.6 (C-4), 163.4 (C-6), 165.6 (C-3), 198.3 (C-8); HRMS: m/z calcd for C<sub>13</sub>H<sub>14</sub>O<sub>6</sub> (M<sup>+</sup>): 266.0790; Found: 266.0762.

2-Naphthalene-2-carboxylic acid (1RS,9RS,11SR,12SR) -12-hydroxy-9-methyl-4-methylene-6,8-dioxo-2,5-dioxatricyclo-[7.3.1.0<sup>3,7</sup>tridec-3(7)-en-11-yl ester (27). Prepared according to general procedure from 22 (43.2 mg, 0.0814 mmol). Purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>–EtOAc, 95:5) to give 27 (9.5 mg, 27.8%) as colorless crystals; mp 226 °C.

IR (KBr) 3474, 3132, 3061, 2924, 1789, 1721, 1589 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 421 (5) [M + H<sup>+</sup>], 251 (100); <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>):  $\delta = 1.30$  (s, 3H, CH<sub>3</sub>), 2.15 (d, 1H, J = 15.1 Hz, 10-H), 2.17 (d, 1H, J = 15.1 Hz, 10-H), 2.31 (dd, 1H, J = 3.9, 16.7 Hz, H-13), 3.35 (d, 1H, J = 16.7 Hz, 13-H), 2.51 (d, 1H, J = 3.0 Hz, OH), 4.50 (q, 1H, J = 3.0 Hz, 12-H), 5.07 (m, 1H, 11-H), 5.17 (m, 1H, 1-H), 5.31 (d, 1H, J=2.8 Hz, CH<sub>2</sub>), 5.42 (d, 1H, J = 2.8 Hz, CH<sub>2</sub>), 7.57 (ddd, 1H, J = 1.3, 7.5, 8.2 Hz,  $H_{arom}$ ), 7.62 (ddd, 1H, J = 1.3, 7.5, 8.2 Hz,  $H_{arom}$ ), 7.90 (d, 2H, J=8.7 Hz,  $H_{arom}$ ), 7.97 (d, 1H, J=8.2 Hz,  $H_{arom}$ ), 7.99 (dd, 1H, J = 1.7, 8.7 Hz,  $H_{arom}$ ), 8.57 (br. s, 1H,  $H_{arom}$ ); <sup>13</sup>C NMR (CD<sub>2</sub>Cl<sub>2</sub>):  $\delta = 26.7$  (CH<sub>3</sub>), 29.7 (C-13), 36.1 (C-10), 48.2 (C-9), 69.1 (C-12), 69.9 (C-11), 83.2 (C-1), 97.2 (CH<sub>2</sub>), 102.7 (C-7), 125.0, 126.8, 127.0, 128.4, 128.7, 129.3, 131.2, 132.4, 135.8 (C<sub>arom</sub>), 148.5 (C-4), 165.6 [CO<sub>2</sub>(C-11)], 166.0 (C-3), 197.6 (C-8); HRMS (DEI+): m/z calcd for  $C_{24}H_{20}O_7$ : 420.1209; Found: 420.1208.

2-Naphthalene-2-carboxylic acid (1RS,9RS,11SR,12SR) -11-hydroxy-9-methyl-4-methylene-6,8-dioxo-2,5-dioxatricyclo-[7.3.1.0<sup>3,7</sup>tridec-3(7)-en-12-yl ester (28). Prepared according to general procedure from 23 (12.0 mg, 0.023 mmol). Purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>–EtOAc, 80:20) to give 28 (3.8 mg, 40.0%) as colorless crystals; mp > 260 °C (decomp.).

IR (KBr) 3428, 3055, 2924, 2853, 1787, 1719, 1669, 1591 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 421 (100)  $[M+H^+]$ ; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 1.38$  (s, 3H, CH<sub>3</sub>), 1.97 (t, 1H, J=12.0 Hz, 10-H), 2.25 (dd, 1H, J=4.7, 16.7 Hz, 13-H), 2.30 (br. d, 1H, J = 12.0 Hz, 10-H); 2.45 (br. d, 1H, J=16.7 Hz, 13-H), 3.99 (br. d, 1H,  $J=9.5 \,\mathrm{Hz}$ , 11-H), 5.29 (m, 1H, 1-H), 5.34 (d, 1H, J = 2.9 Hz, C=CH<sub>2</sub>), 5.44 (d, 1H, J = 2.9 Hz, C=CH<sub>2</sub>), 5.66 (m, 1H, 12-H), 7.61 (d, 1H, J = 8.1 Hz,  $H_{arom}$ ), 7.64 (dd, 1H, J=1.4, 8.0 Hz,  $H_{arom}$ ), 7.90–7.95 (m, 2H,  $H_{arom}$ ), 8.00 (d, 1H, J=8.1 Hz,  $H_{arom}$ ), 8.03 (dd, 1H, J = 1.7, 8.6 Hz, H<sub>arom</sub>), 8.60 (s, 1H, H<sub>arom</sub>); <sup>13</sup>C NMR  $(100 \text{ MHz}; \text{CDCl}_3): \delta = 26.9 \text{ (CH}_3), 30.6 \text{ (C-13)}, 40.7 \text{ (C-13)}$ 10), 47.9 (C-9), 65.2 (C-11), 71.8 (C-12), 80.6 (C-1), 97.6 (CH<sub>2</sub>), 102.5 (C-7), 124.9, 125.0, 127.1, 127.9, 128.7, 128.7, 129.0, 129.4, 129.5, 131.7 (C<sub>arom</sub>), 147.8 (C-4),

165.2 (C-6), 165.9 [ $CO_2$ (C-12)], 174.0 (C-3), 197.1 (C-8); HRMS (EI) m/z calcd for  $C_{24}H_{20}O_7$ : 420.1209; Found: 420.1161.

2-Naphthalene-2-carboxylic acid (1RS,9RS,11SR,12SR) -12-(naphthalene-2-carbonyloxy)-9-methyl-4-methylene-6,8-dioxo-2,5-dioxatricyclo-[7.3.1.0<sup>3,7</sup>tridec-3(7)-en-11-yl ester (29). Prepared according to general procedure from 23 (40.6 mg, 0.059 mmol). Purified by flash chromatography (*n*-hexane-CH<sub>2</sub>Cl<sub>2</sub>-EtOAc, 50:40:10) to give 29 (6.4 mg, 18.8%) as colorless crystals; mp 255–260 °C (decomp.).

IR (KBr) 3132, 3055, 2924, 2847, 1786, 1725, 1669, 1583 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub><sup>+</sup>): m/z (%) = 575 (34)  $[M+H^+]$ , 235 (100); <sup>1</sup>H NMR: (CDCl<sub>3</sub>):  $\delta = 1.47$  (s, 3H, CH<sub>3</sub>), 2.31 (t, 1H, J = 12.8 Hz, 10-H), 2.41 (dd, 1H, J=4.6, 16.7 Hz, 13-H), 2.47 (dd, 1H, J=4.4, 12.8 Hz, 10-H), 2.58 (d, 1H, J = 16.7 Hz, 13-H), 5.34 (m, 1H, 11-H), 5.36 (m, 1H, 1-H), 5.40 (d, 1H, J = 3.0 Hz,  $C = CH_2$ ), 5.50 (d, 1H,  $J = 3.0 \,\text{Hz}$ ,  $C \doteqdot CH_2$ ), 6.00 (m, 1H, 12-H), 7.43 (t, 1H,  $J = 7.5 \,\text{Hz}$ ,  $H_{\text{arom}}$ ), 7.54 (t, 1H,  $J = 7.5 \,\text{Hz}$ ,  $H_{arom}$ ), 7.61 (t, 1H,  $J = 7.5 \,\text{Hz}$ ,  $H_{arom}$ ), 7.64–7.69 (m, 2H,  $H_{arom}$ ), 7.76 (d, 1H, J = 8.8 Hz,  $H_{arom}$ ), 7.80 (d, 1H,  $J = 8.3 \text{ Hz}, \text{ H}_{arom}$ ), 7.87 (dd, 1H,  $J = 1.7, 8.7 \text{ Hz}, \text{ H}_{arom}$ ), 7.94 (d, 1H, J = 8.3 Hz,  $H_{arom}$ ), 7.96 (d, 1H, J = 8.8 Hz,  $H_{arom}$ ), 7.98 (d, 1H,  $J=8.3 \, Hz$ ,  $H_{arom}$ ), 8.05 (dd, 1H,  $J = 1.7, 8.7 \text{ Hz}, H_{arom}$ ), 8.38 (s, 1H,  $H_{arom}$ ), 8.62 (s, 1H,  $H_{arom}$ ); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 27.1$  (CH<sub>3</sub>), 30.7 (C-13), 37.4 (C-10), 47.8 (C-9), 66.9 (C-11), 69.2 (C-12), 80.4 (C-1), 97.9 ( $C = CH_2$ ), 102.6 (C-7), 125.2, 125.3, 126.4, 126.7, 127.0, 127.4, 128.0, 128.2, 128.6, 128.8, 129.0, 129.3, 129.6, 129.8, 131.7, 132.0, 132.6, 132.8, 135.9, 136.2 (C<sub>arom</sub>), 147.7 (C-4), 165.2 (C-6), 165.2, 165.3  $[CO_2(C-11), CO_2(C-12)], 196.9 (C-8). HRMS (EI): m/z$ calcd for C<sub>35</sub>H<sub>26</sub>O<sub>8</sub>: 574.1637; Found: 574.1628.

(1RS,9RS,11RS,12RS) - 11,12 - Dihydroxy - 9 - methyl - 4-methylene - 2,5 - dioxatricyclo - [7.3.1.0 $^{3,7}$ tridec - 3(7) - ene-6,8-dione (26). Prepared according to general procedure from 20 (6.4 mg, 0.017 mmol). Purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>-EtOAc—acetone–*i*-PrOH, 45:40:10:5) to give 26 (1.0 mg, 22.2%) as colorless crystals; mp > 280 °C (decomp.).

IR (KBr) 3441, 2924, 1778, 1648, 1594 cm<sup>-1</sup>; MS (CI, CH<sub>5</sub>+): m/z (%) = 267 (62) [M+H+], 242 (100);  $^{1}$ H NMR (acetone- $d_6$ ):  $\delta$  = 1.11 (s, 3H, CH<sub>3</sub>), 1.74 (d, 1H, J=14.2 Hz, CH<sub>2</sub>), 2.13 (dd, 1H, J=4.0, 16.6 Hz, CH<sub>2</sub>), 2.31 (dt, 1H, J=3.5, 14.2 Hz, CH<sub>2</sub>), 2.68 (dt, 1H, J=3.0, 16.2 Hz, CH<sub>2</sub>), 3.84 (m, 1H, 12-H), 3.99 (m, 1H, 11-H), 4.08 (d, 1H, J=3.0 Hz, OH), 4.15 (d, 1H, J=9.0 Hz, OH), 5.13 (d, 1H, J=2.6 Hz, C=CH<sub>2</sub>), 5.15 (m, 1H, 1-H), 5.23 (d, 1H, J=2.6 Hz, C=CH<sub>2</sub>);  $^{13}$ C NMR (acetone- $d_6$ ):  $\delta$ =27.1 (CH<sub>3</sub>), 34.2 (C-13), 44.0 (C-10), 45.4 (C-9), 68.8 (C-12), 72.0 (C-11), 83.2 (C-1), 94.4 (=CH<sub>2</sub>), 101.1 (C-7), 150.1 (C-4), 161.4 (C-6), 166.4 (C-3), 199.0 (C-8); HRMS (EI): m/z calcd for C<sub>13</sub>H<sub>14</sub>O<sub>6</sub>: 266.0790; Found 266.0825.

(1*RS*,9*RS*)-9-Methyl-4-methylene-2,5-dioxatricyclo[7.3.1.O <sup>3,7</sup>]trideca-3(7),11-diene-6,8-dione (6). Prepared according to general procedure from **18** (61.9 mg, 0.18 mmol).

Purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>–EtOAc, 97:3) to give 6 (21.4 mg, 50.9%) as colorless crystals; mp > 300 °C (decomp.).

IR (KBr) 3041, 2875, 1790, 1672, 1652, 1589 cm<sup>-1</sup>; MS (CI; CH<sub>5</sub><sup>+</sup>): m/z = 233 (100) [M+H<sup>+</sup>]; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 1.22$  (s, 3H, CH<sub>3</sub>), 1.93 (dq, 1H, J = 2.3, 18.0 Hz, 10-H), 2.13 (dd, 1H, J = 6.4, 16.4 Hz, 13-H), 2.47 (d, 1H, J = 16.4 Hz, 13-H), 2.71 (dd, 1H, J = 6.1, 18.0 Hz, 10-H), 5.23 (d, 1H, J = 2.6 Hz, C=CH<sub>2</sub>), 5.30 (d, 1H, J = 2.6 Hz, C=CH<sub>2</sub>), 5.34 (m, 1H, 1-H), 5.75 (d<sub>br</sub>, 1H, J = 10.0 Hz, 12-H), 6.18 (ddt, 1H, J = 1.6, 6.1, 10.0 Hz, 11-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 25.9$  (CH<sub>3</sub>), 34.4 (C-13), 37.4 (C-10), 44.8 (C-9), 77.0 (C-1), 96.2 (C=CH<sub>2</sub>), 102.6 (C-7), 121.5 (C-12), 135.2 (C-11), 148.2 (C-4), 164.2 (C-3), 164.5 (C-6), 198.6 (C-8); HRMS (EI): m/z calcd for C<sub>13</sub>H<sub>12</sub>O<sub>4</sub>: 232.0733; Found: 232.0736.

#### Antibacterial activities

The compounds were evaluated in vitro for their antimicrobial activities against Gram-positive [S. aureus ATCC 25923, S. aureus ATCC 29213 (MRSA) and E. faecalis ATCC 29212] bacteria at concentrations ranging from 0.5 to 128 µg/mL. Minimum inhibitory concentration (MIC) values were determined by a broth microdilution method carried out according to DIN 58940-8.16 Each compound was dissolved in dimethyl sulfoxide to give a 2.56 mg/mL stock solution, which was serially diluted 2-fold with Mueller-Hinton broth. Overnight cultures of the test organisms were suspended in saline, and the cell density was adjusted to give an initial inoculum of 10<sup>6</sup> colony forming units per milliliter (CFU mL<sup>-1</sup>) in Mueller-Hinton broth. To each spot of a 96-well micro-plates containing 100 μL of graded concentrations of each compound 100 µL of the bacterial suspension of (10<sup>6</sup> CFU mL<sup>-1</sup>) was added to yield a final inoculum of  $5 \times 10^5$  CFU mL<sup>-1</sup>. After incubation at 37 °C for 18 h the MIC was defined as the minimum concentration of a test compound that resulted in no visible growth of bacteria, compared with the drug-free control. It should be noted that the concentration of dimethyl sulfoxide in the medium did not affect the growth of any of the microorganisms tested. The MIC of each compound was determined at least twice.

### Cytotoxicity

The cytotoxicity of compounds 5–7 and 27 was determined using two cell lines: human acute myeolid leukemia HL-60 cells (DSMZ ACC 3) and human T cell leukemia Jurkat cells (DSMZ ACC 282). Impaired cell viability was measured using the MTT assay according to Mosmann, based on the ability of viable cells to reduce yellow MTT to blue formazan. Solutions of the test compounds were freshly prepared from stock solutions in dimethyl sulfoxide, which were diluted to the required concentrations. Final DMSO concentrations were 1% (v/v). The test was carried out in 96-well plates with an inoculum of  $9\times10^5$  cells mL<sup>-1</sup>. Total assay volume was  $101\,\mu$ L. The cells were exposed to the test

compounds at 37 °C for 24 h. Then they were incubated with methylthiazolyldiphenyl tetrazolium chloride (MTT, 0.5 mg/mL, 37 °C, 2 h) and subsequently solubilized in DMSO (200 μL) for at least 2 h in the dark. The extend of reduction of MTT was quantified by absorbance measurement (A<sup>550 nm</sup>, SLt Spectra, SLT Labinstruments, Crailsheim, Germany). For determination of the IC<sub>50</sub> values, the optical density was plotted against the log concentration. At least four different concentrations have been tested. Every test was performed at least in triplicates, and all experiments have been repeated at least twice. Positive control measurements were performed with actinomycin.

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#### References and Notes

- 1. Johnson, A. P.; Woodford, N. J. Antimicrob. Chemother. **2002**, 50, 621 and references cited therein.
- 2. Vincent, J.-L. Intensive Care Med. 2000, 26, 83.
- 3. (a) Isolation and biological evaluation: Tsuchida, T.; Iinuma, H.; Nishida, C.; Kinoshita, N.; Sawa, T.; Hamada, M.; Takeuchi, T. *J. Antibiot.* **1995**, *48*, 1104. (b) Structure determination: Tsuchida, T.; Iinuma, H.; Sawa, R.; Takahashi, Y.; Nakamura, H.; Nakamura, K. T.; Sawa, T.; Naganawa, H.; Takeuchi, T. *J. Antibiot.* **1995**, *48*, 1110.
- 4. Tsuchida, T.; Iinuma, H.; Nakamura, K. T.; Nakamura, H.; Sawa, T.; Hamada, M.; Takeuchi, T. *J. Antibiot.* **1995**, *48*, 1330.
- 5. A part of this work has been communicated in preliminary form: Paintner, F. F.; Allmendinger, L.; Bauschke, G.; Polborn, K. *Synlett* **2002**, 1308.
- 6. Paintner, F. F.; Allmendinger, L.; Bauschke, G. Synthesis 2001, 2113.
- 7. Paintner, F. F.; Bauschke, G.; Kestel, M. Tetrahedron Lett. 2000, 41, 9977.
- 8. Stork, G.; Logusch, E. W. Tetrahedron Lett. 1979, 3361.
- 9. For a mechanistic discussion of this key ring closure step, see: ref 5.
- 10. Shing, T. K. M.; Tam, E. K. W.; Tai, V. W.-F.; Chung, I. H. F.; Jiang, Q. Chem. Eur. J. 1996, 2, 50.
- 11. VanRheenen, V.; Kelly, R. C.; Cha, D. Y. *Tetrahedron Lett.* **1976**, 1973.
- 12. Mukaiyama, T.; Tabusa, F.; Suzuki, K. Chem. Lett. 1983,
- 13. (a) The facile oxidation of sulfides to sulfones in the presence of C–C double bonds using catalytic osmium tetroxide and *N*-methylmorpholine-*N*-oxide or trimethylamine-*N*-oxide as co-oxidant has been reported: Kaldor, S. W.; Hammond, M. *Tetrahedron Lett.* **1991**, *32*, 5043. (b) Priebe, W.; Grynkiewicz, G. *Tetrahedron Lett.* **1991**, *32*, 7353.
- 14. (a) On the other hand, sulfides are known to be essentially inert to oxidation by osmium tetroxide under stoichiometric conditions: Stork, G.; van Tamelen, E. E.; Friedman, L. J.; Burgstahler, A. W. J. Am. Chem. Soc. 1953, 75, 384. (b) Djerassi, C.; Engle, R. R. J. Am. Chem. Soc. 1953, 75, 3838. (c)

Henbest, H. B.; Khan, S. A. *J. Chem. Soc., Chem. Commun.* **1968**, 1036. (d) For chemoselective oxidations of C–C double bonds in the presence of sulfides with catalytic OsO<sub>4</sub> and K<sub>3</sub>Fe(CN)<sub>6</sub> as co-oxidant, see: Walsh, P. J.; Ho, P. T.; King, B.; Sharpless, K. B. *Tetrahedron Lett.* **1994**, *35*, 5129.

- 15. Pollet, P.; Gelin, S. Tetrahedron 1978, 34, 1453.
- 16. Methods for Susceptibility Testing of Bacterial Pathogens (except mycobacteria) against Chemotherapeutic Agents (DIN paperback); Beuth: Berlin, Vienna, Zurich, 2000; Vol. 222.
- 17. Mosmann, T. J. Immunol. Methods 1983, 65, 55.